ACYL COENZYME A INHIBITION OF Leuconostoc mesenteroides
GLUCOSE-6-PHOSPHATE DEHYDROGENASE: A COMPARISON OF THE
TPN AND DPN LINKED REACTIONS

Elmon L. Coe and Lung-Hsiung Hsu

Department of Biochemistry
Northwestern University Medical School
Chicago, Illinois 60611

Received May 18, 1973

The inhibitory effects of ATP, coenzyme A, and acetyl, malonyl, and oleyl derivatives of coenzyme A on the TPN and DPN dependent activities of Leuconostoc glucose-6-phosphate dehydrogenase are compared. At pH 7.8, 24°, saturating levels of DPN or TPN, and inhibitor concentrations of 2-4 mM only ATP has an appreciable effect on the TPN dependent reaction, but all were potent inhibitors of the DPN dependent reaction. Oleyl coenzyme A was the most effective (K_i ~ 0.15 mM against glucose-6-phosphate) while acetyl coenzyme A was least effective (K_i ~ 1.0 mM). A possible regulatory role of this inhibition in fatty acid synthesis is suggested.

Glucose-6-phosphate dehydrogenase (EC 1.1.1.49) from Leuconostoc mesenteroides is known to function with both TPN and DPN as hydrogen acceptors. The isotope distribution studies, Kemp and Rose deduced that TPNH and DPNH generated by this enzyme are reoxidized via distinctly different pathways, the DPNH donating its hydrogen to the end products of fermentation, and the TPNH providing hydrogen for reductive syntheses, particularly fatty acid synthesis. In 1967, Olive and Levy crystallized the enzyme and from comparative inhibition studies concluded that the reactions with TPN and DPN were both catalyzed by a single enzyme and that both coenzymes are bound to the same site. More recently, Hsu observed that both reactions were inhibited by ATP; this inhibition is competitive with respect to glucose-6-phosphate, but the inhibition of the DPN-linked reaction is the more severe, the Ki being 0.3-0.5 mM compared to 1.5-2.0 mM for the TPN-linked reaction. This suggested that an increasing ATP level would tend to shift the hydrogens generated by this enzyme avay from reduction of fermentation products toward reductive syntheses.

A possible complementary control mechanism was suggested by the observation that long-chain acv1 coenzyme A derivatives inhibited the TPNlinked glucose-6-phosphate dehydrogenase activities of yeast, rat liver, rat adipose tissue, and human erythrocytes. 5,6 This inhibition was also competitive with respect to glucose-6-phosphate and was evident at very low concentrations of the coenzyme A derivatives. With the yeast enzyme, the K; values for the palmity16 and steary15 derivatives were 0.003 and 0.004 mM. respectively. Hence, it appeared possible that acyl-coenzyme A could "turn off" the TPNH generating function of the Leuconostoc enzyme and thereby act in opposition to the ATP effect. To test this possibility the Leuconostoc enzyme was assayed in the presence of several coenzyme A derivatives. A comparison of the effect on the TPN-linked and DPN-linked activities are shown in Table I, and it may be seen that, contrary to expectations, coenzyme A and acyl coenzyme A exert their major effect against the DPN-linked system. In fact, ATP appears to be the most effective inhibitor of the TPN-linked system. The approximate Ki values (DPN system only) are calculated on the assumption that the inhibition is competitive with respect to glucose-6phosphate; the data are consistent with this assumption, although in some instances they are not sufficiently precise to exclude other types of inhibition. From these approximations, however, it is apparent that even the long chain oleyl-coenzyme A is much less effective against the Leuconostoc enzyme than the palmityl or stearyl-coenzyme A is against the yeast or mammalian dehydrogenases. It is also apparent that the oleyl-coenzyme A is the most effective inhibitor for the DPN dependent reaction.

Although the original hypothesis that TPNH generation by the dehydrogenase is selectively turned off by acyl-coenzyme A is shown to be invalid by these results, an alternate possibility for regulation is suggested by the observation of Ilton et al.⁷ that the presence of DPNH substantially decreases the TPNH level required for optimal activity of the fatty acid synthetase from Mycobacterium phlei. If the systems from Leuconostoc and Mycobacterium are

TABLE I

Relative Inhibitions of TPN and DPN Dependent Glucose-6-Phosphate Dehydrogenase Activities by ATP and Coenzyme A Derivatives

INHIBITOR	Conc. mM	% Inl TPN	hibition DPN	Approx. K; (with DPN)
ATP	3.9	50	90	0.5
CoA	3.4	12	82	0.7
AcCoA	2.6	0	70	1.0
Ma1CoA	2.4	0	86	0.4
Oleyl CoA	2.0	9	90	0.15

CONDITIONS USED FOR ASSAY:

REACTION MIXTURE: 40 mM tris buffer, pH 7.8; 0.4-0.5 mM DPN or 0.2 mM TPN; 0.02-0.2 mM glucose-6-phosphate; 0.07 µ/ml glucose-6-phosphate dehydrogenase; 23-24°; concentrations of ATP, coenzyme A derivatives as indicated. Absorption increase at 340 nm followed with a Gilford recording spectrophotometer. SOURCES: Leuconostoc mesenteroides enzyme, Worthington Biochemical Corp., Freehold, New Jersey; ATP, glucose-6-phosphate, and Tris, Sigma Chemical Co., St. Louis, Mo.; coenzyme A and acyl-coenzyme A derivatives, P-L Biochemicals, Inc., Milwaukee, Wisconsin.

KINETIC PARAMETERS OF Leuconostoc ENZYME: Km for TPN, 0.014 mM; for DPN, 0.13 mM; for glucose-6-phosphate, 0.17 mM with either coenzyme.

comparable in this regard, then accumulation of long chain acyl-coenzyme A derivatives might indirectly curtail fatty acid synthesis by decreasing the DPNH level, thereby lowering the rate of reaction with TPNH. Such a mechanism would be more selective than a simple suppression of TPNH generation in that it could redirect the continuing supply of TPNH into other biosynthetic pathways.

ACKNOWLEDGEMENTS

The authors wish to thank Mrs. Jaya Subbarao for her technical assistance; this work was supported by grant no. CA-10723 of the National Institutes of Health, U.S.A.

REFERENCES

 DeMoss, R. D., Gunsalus, I. C., and Bard, R.C., J. Bacteriol., 66, 10 (1953).

- 2. Kemp. C. R. and Rose, A. I., J. Biol. Chem., 239, 2998 (1964).
- 3. Olive, C. and Levy, H. R., Biochem., 6, 730 (1967).
- 4. Hsu, L. H., Studies on Glucose-6-Phosphate Dehydrogenase from Ascites Tumor Cells, Slime Mold, and L. mesenteroides. Ph.D. Dissertation, Northwestern University, Evanston, Illinois; June, 1971.
- 5. Eger-Neufeldt, F., Teinzer, A., Weiss, L., and Wieland, O. Biochem. Biophys. Res. Commun., 19, 43 (1965).
- 6. Taketa, K. and Pogell, B. M., J. Biol. Chem., 241, 720 (1966).
- 7. Ilton, M., Jevans, A. W., McCarthy, E. D., Vance, D., White, H. B. and Block, K., Proc. Nat. Acad. Sci., U.S., 68 87 (1971).